

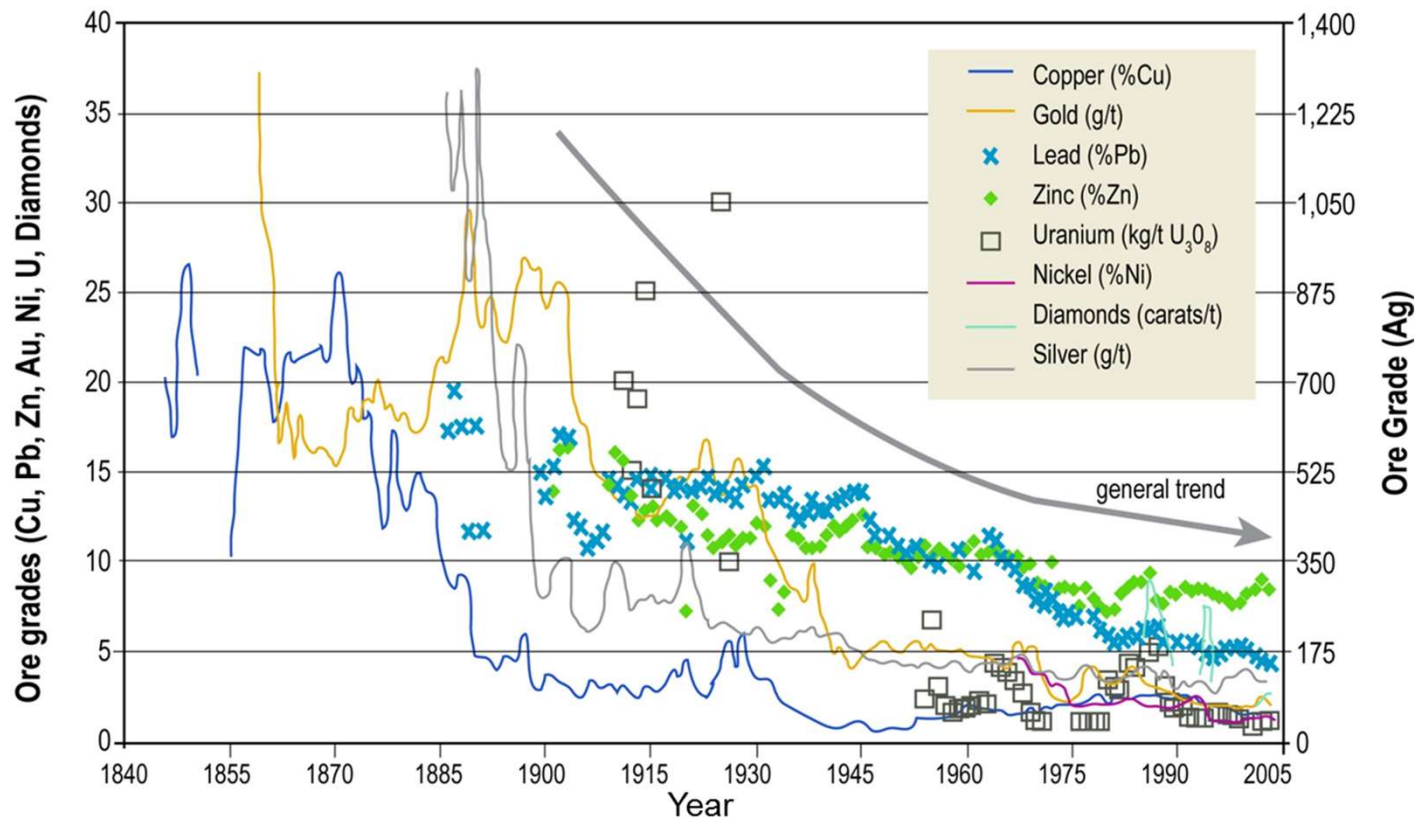
Potential applications of biomining for in situ recovery

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Declining ore grades



→ Motivation for biomining and *in situ* recovery

Principles of biomining and bioleaching

- **Biomining:** the use of microorganisms for extraction of metals from minerals and wastes and recovery of metals from leach liquors
 - **Bioleaching:** the use of microorganisms for the extraction of metals from minerals and wastes through redoxolysis, acidolysis and/or complexolysis
 - **Biorecovery:** the use of microorganisms for the recovering metals from leach liquors through bioprecipitation and/or biosorption

Bioleaching through redoxolysis

Biological process	Description	Electron donors and acceptors
Oxidative bioleaching	Fe ²⁺ biooxidation for oxidative ferric leaching of sulfide minerals	Electron donor: Fe ²⁺ Electron acceptors: O ₂ , NO ₃ ⁻
	Mn ²⁺ oxidation to Mn ⁴⁺ for use of MnO ₂ as an oxidant for bioleaching sulfide minerals	Electron donor: Mn ²⁺ Electron acceptor: O ₂
Reductive bioleaching	Fe ³⁺ bioreduction for reductive bioleaching of oxide minerals	Electron donors: organic compounds, H ₂ , reduced sulfur compounds Electron acceptor: Fe ³⁺
	Mn ⁴⁺ bioreduction for reductive bioleaching of Mn ores	Electron donors: organic compounds Electron acceptor: Mn ⁴⁺

Bioleaching through acidolysis

Biological process	Description	Electron donors and acceptors
Acidolysis with inorganic acid	Sulfur biooxidation to sulfuric acid for leaching acid-soluble minerals and wastes	Electron donors: reduced sulfur compounds Electron acceptors: O_2 , Fe^{3+}
Acidolysis with organic acids	Biogenic organic acids production for leaching acid-soluble minerals and wastes	Electron donor: organic compounds Electron acceptor: O_2 , none (fermentation)

Bioleaching through complexolysis

Biological process	Description	Electron donors and acceptors
Complexolysis	Biogenic organic acids production for rare earth element leaching	<p>Electron donors: organic compounds</p> <p>Electron acceptors: Fe^{3+}, O_2, none (fermentation)</p>
	Biogenic cyanide production for gold leaching	<p>Electron donors: organic compounds</p> <p>Electron acceptor: O_2</p>
	Iodide (I^-) biooxidation to iodine (I_2) for gold leaching	<p>Electron donor: I^-</p> <p>Electron acceptor: O_2</p>

Contact, non-contact and collaborative bioleaching of sulfide minerals

Non-contact leaching

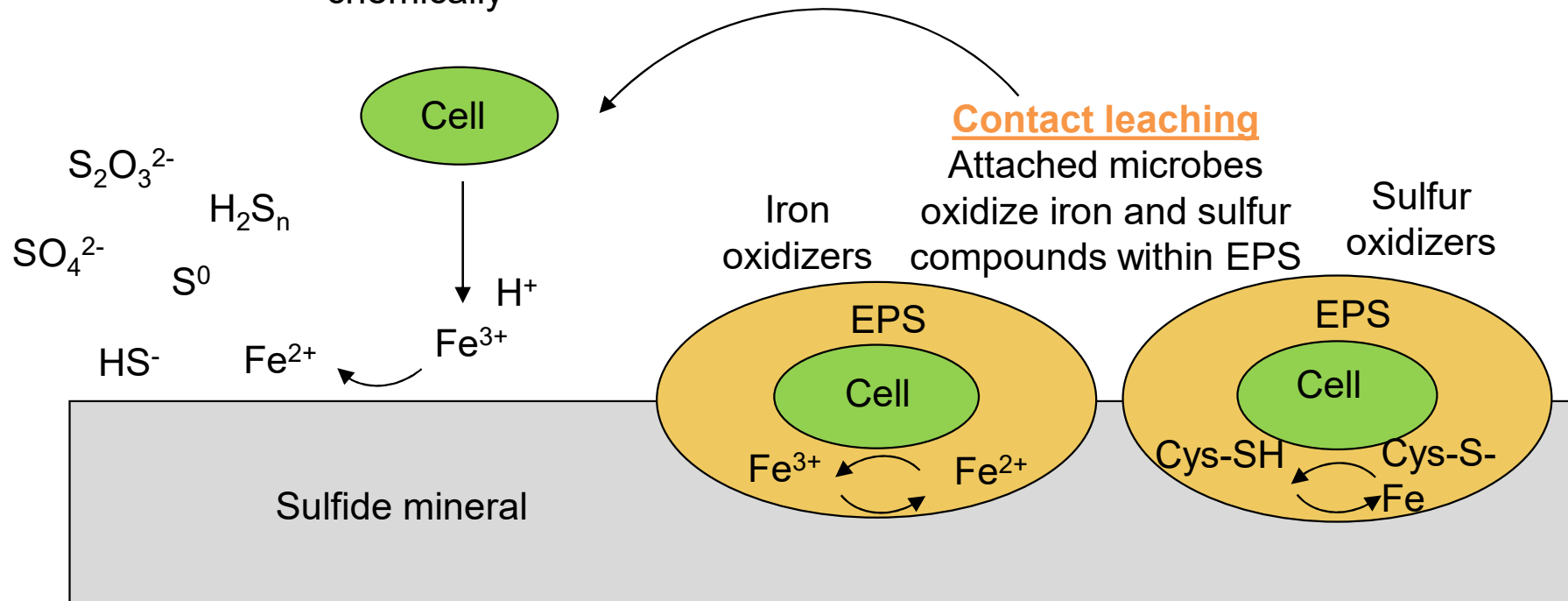
Planktonic microbes produce Fe^{3+} and/or H^+ which solubilize minerals chemically

Collaborative leaching

Attached cells release sulfur colloids for planktonic sulfur oxidizers

Contact leaching

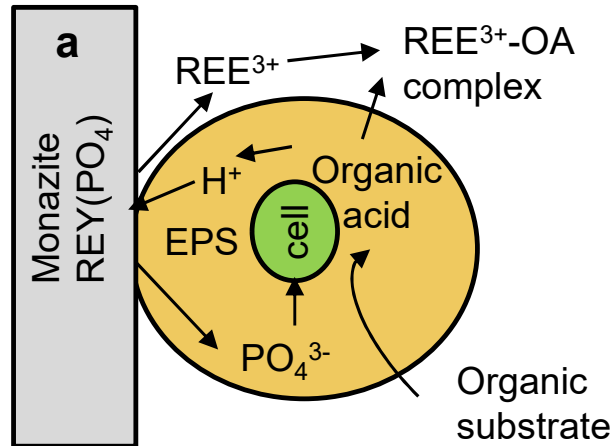
Attached microbes oxidize iron and sulfur compounds within EPS



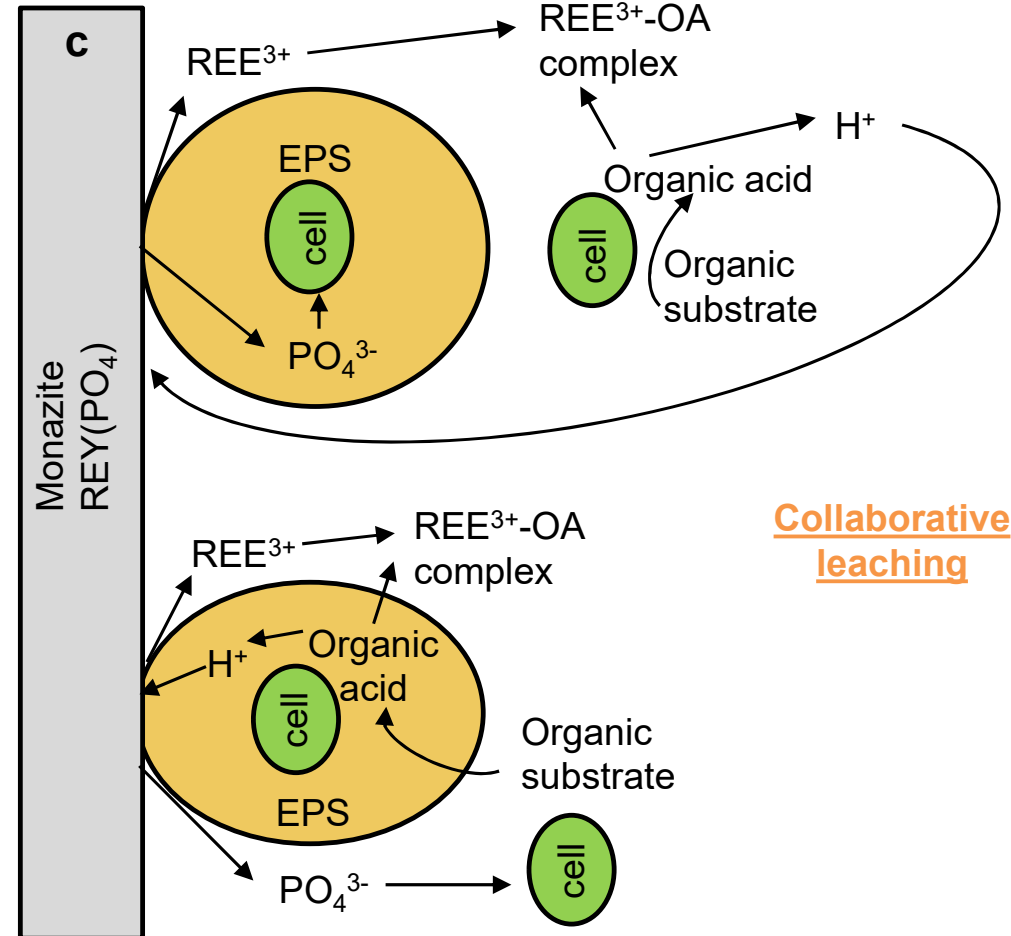
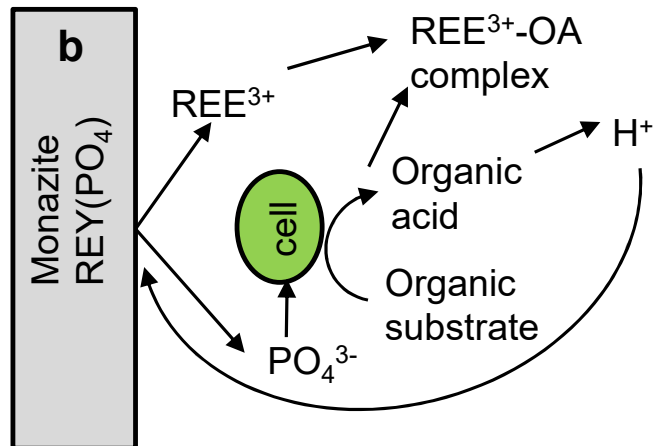
EPS = exopolysaccharides

Contact, non-contact and collaborative bioleaching of rare earths from monazite

Contact leaching



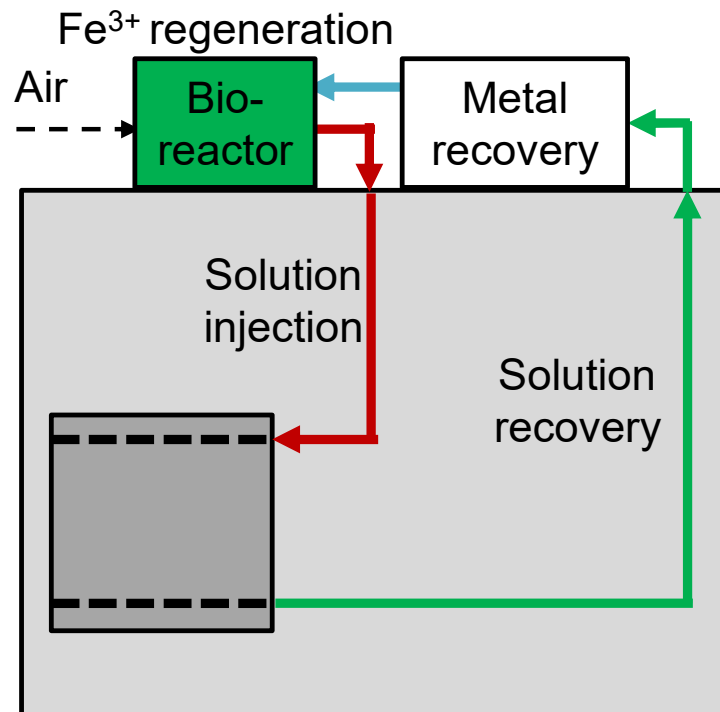
Non-contact leaching



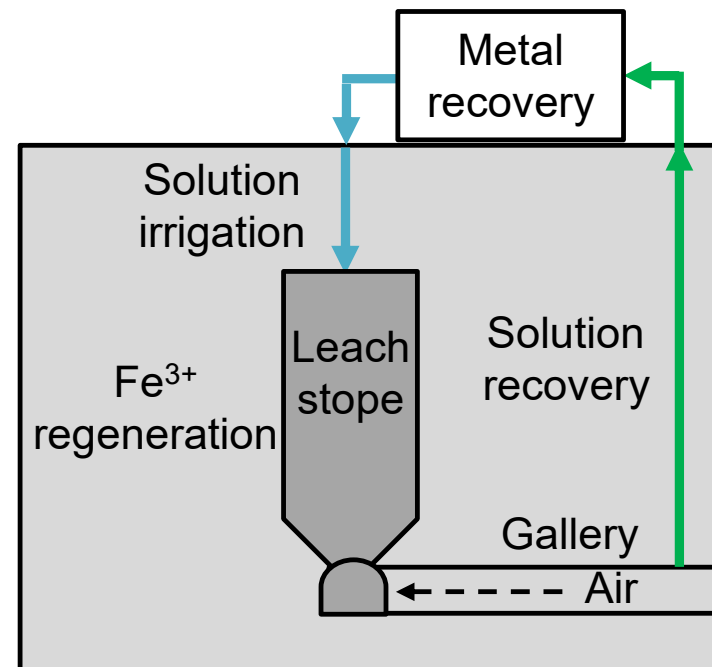
Collaborative leaching

In situ biomining of under saturated and unsaturated conditions

Saturated *in situ* biomining (biogenic lixiviant generation and leaching in separate unit processes)

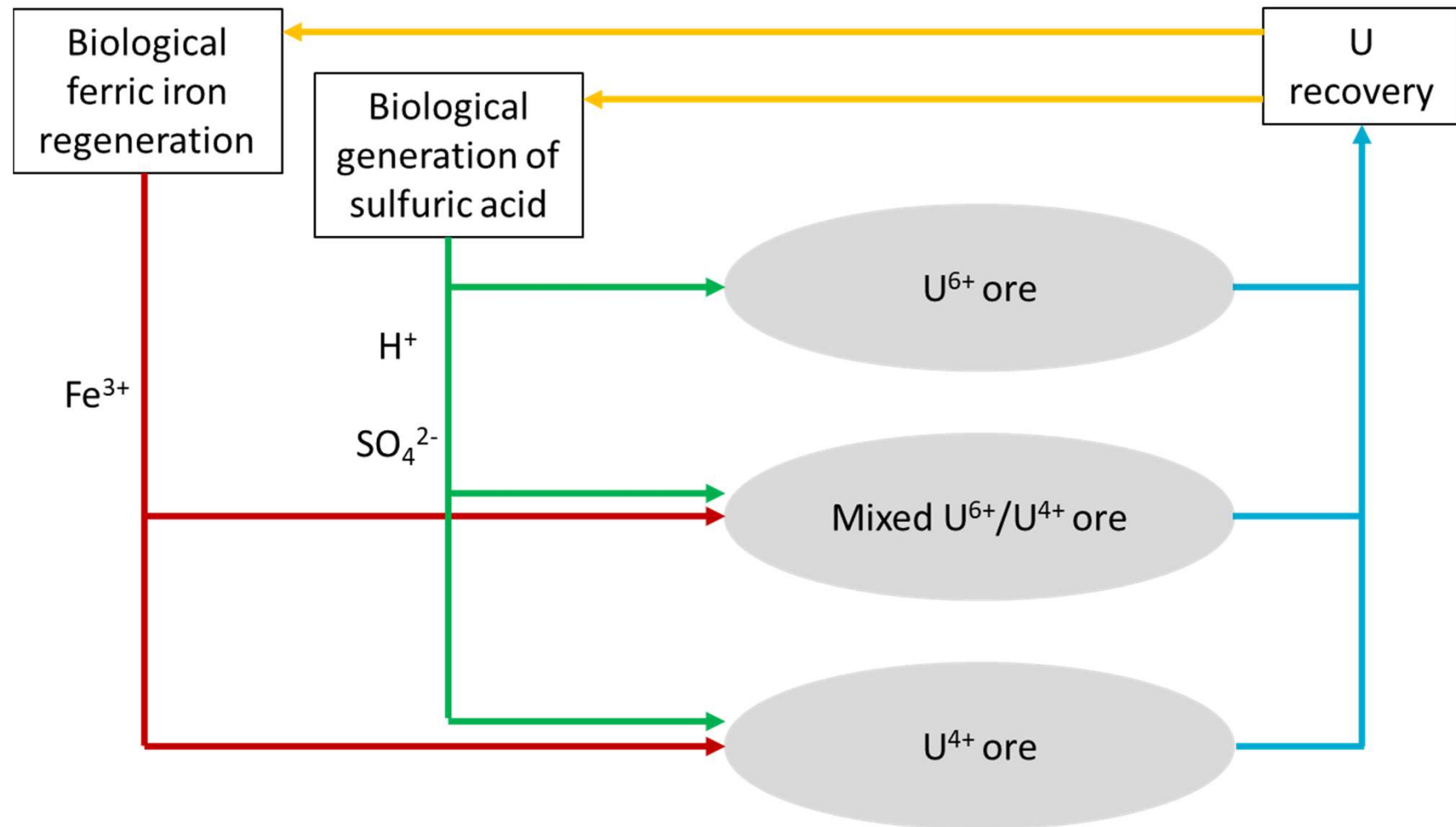


Unsaturated *in situ* biomining (biogenic lixiviant generation and leaching in a single unit process)



Adapted from: Kaksonen and Pedersen 2023. Chapter 17: The Future of Biomining: Towards Sustainability in a Metal-demanding World. In: Bryan, Johnson, Roberto and Schlömann (eds.) *Biomining Technologies: Bioprocessing Options for Extracting and Recovering Metals from Ores and Wastes*. Springer-Verlag (Heidelberg, Germany). Pp. 295-314.

In situ bioleaching of uranium



***In situ* stope bioleaching of uranium (Ontario, Canada)**

■ **Stanrock Mine (1960s)**

- Nutrient addition did not improve leaching

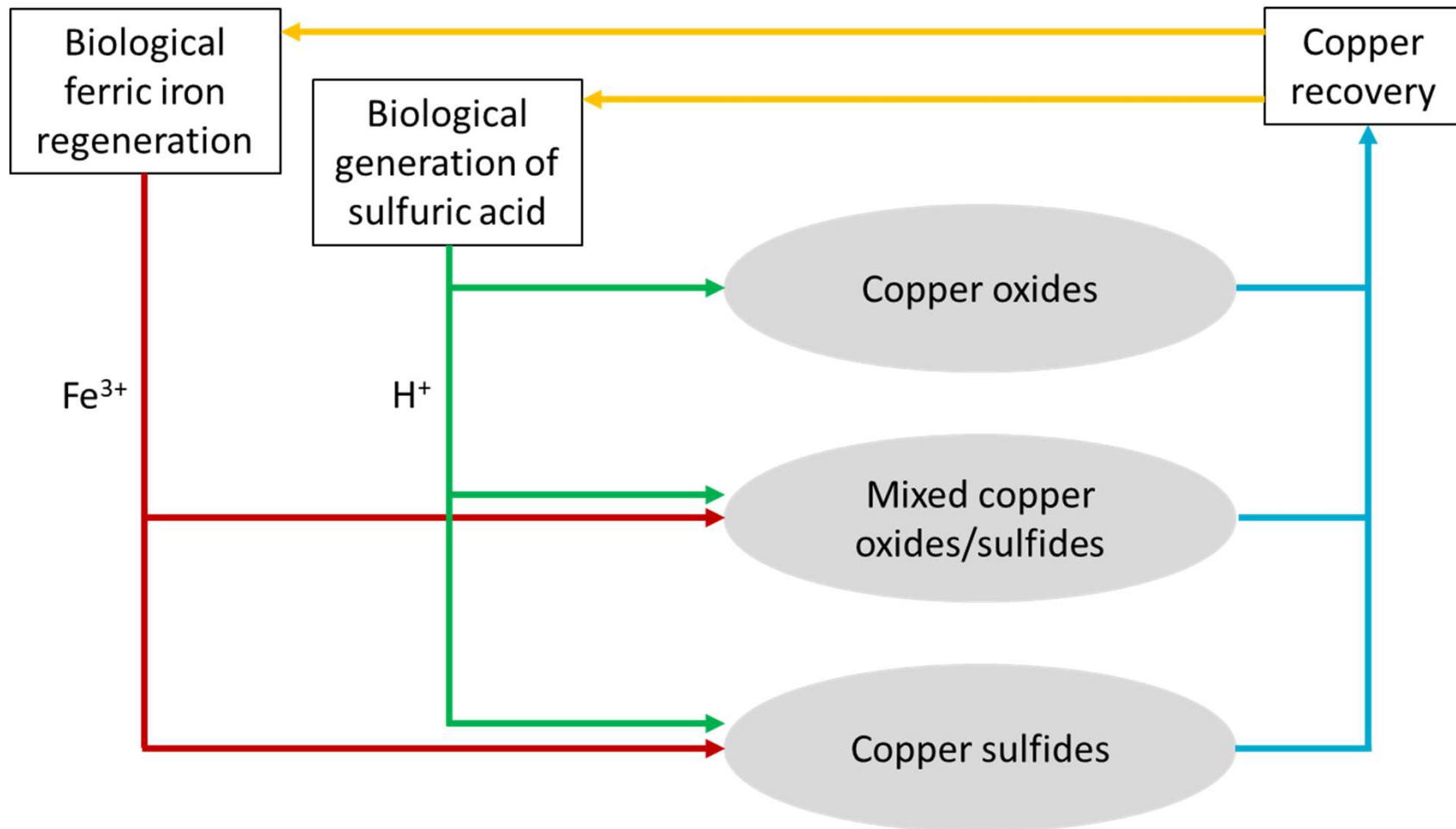
■ **Agnew Lake Mine (late 1970s)**

- Challenges with ore fracturing, and enabling sufficient contact between solution and minerals while containing the leach solution

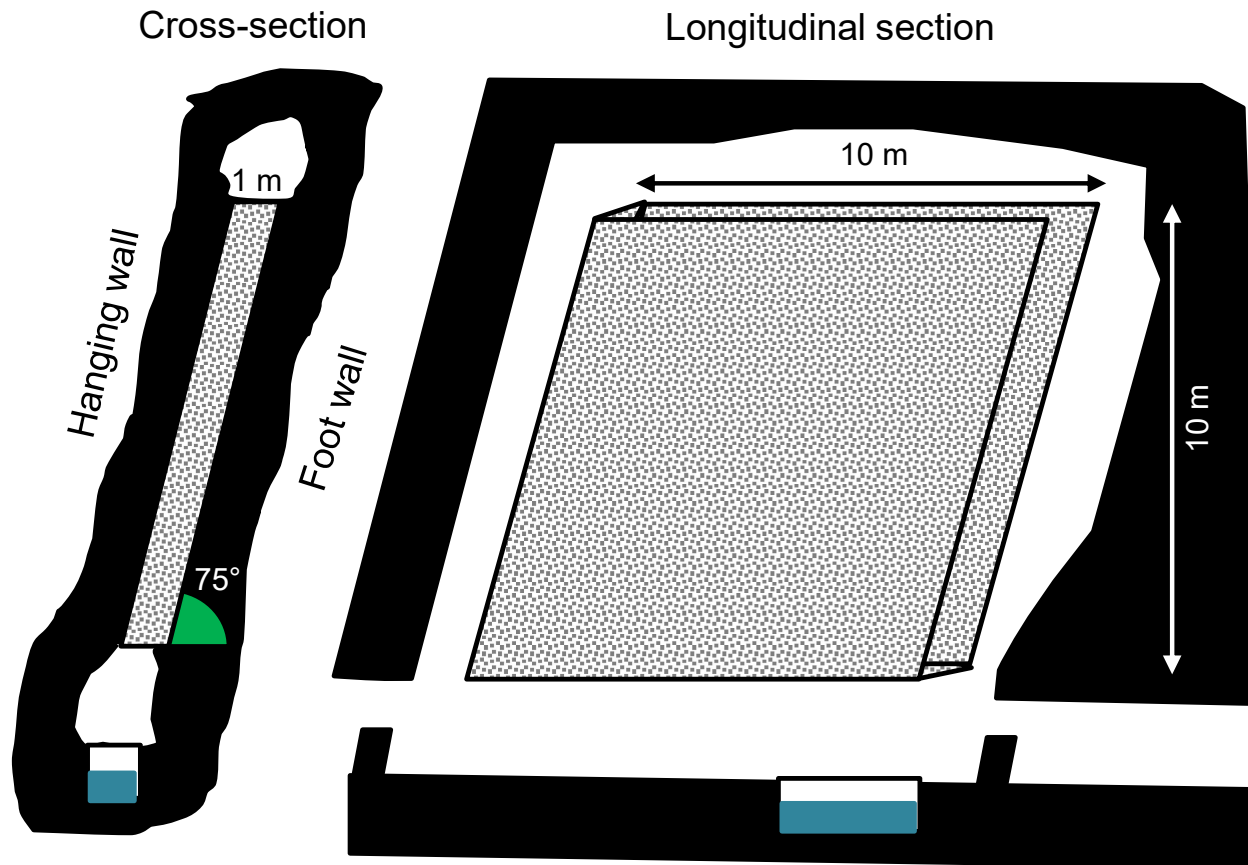
■ **Denison Mine (1980s and early 1990s)**

- Ore was fractured and concrete bulkhead constructed across the opening of a horizontal shaft
- Ore behind bulkhead was flooded with leach liquor in cycles and pregnant leach liquor drained after 3 weeks for U recovery
- Bacteria and acidic ferric sulfate leaching improved yields compared to chemical acid leaching

In situ bioleaching of copper



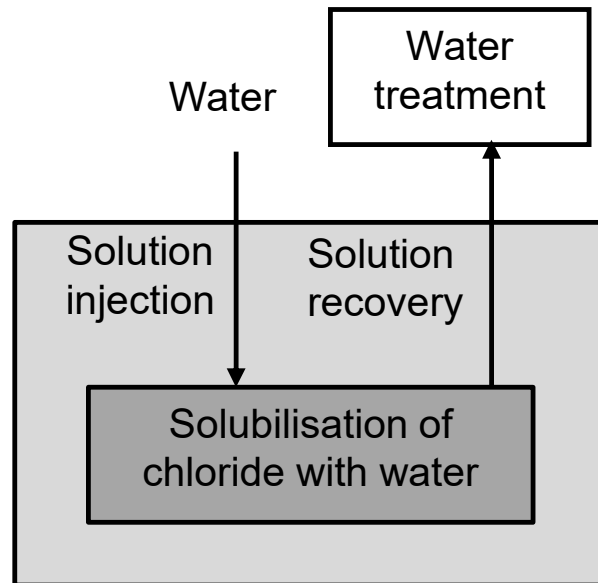
In situ stope bioleaching of Cu and Zn at Ilba Mine (Romania)



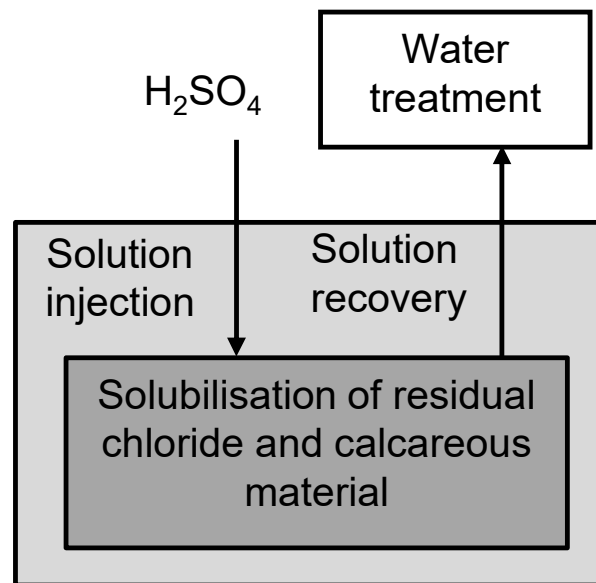
- Sulfidic ore: pyrite (FeS_2), chalcopyrite (CuFeS_2), covellite (CuS), bornite (Cu_5FeS_4), sphalerite (Zn,FeS), galena (PbS), marcasite (FeS_2)
- Ore blasted to <30 cm rocks
- 30% of ore removed \rightarrow 70 m³ ore leached in 10 m x 10 m x 1 m stope
- Inoculation with acidophilic Fe- and S-oxidisers
- Leach liquor aerated and circulated intermittently
- 10% Cu and 78% Zn leaching after 18 months
- Challenges with ore humidification and in winter access to energy for aeration and liquor circulation

In situ biominining of copper from a saline calcareous copper sulfide ore

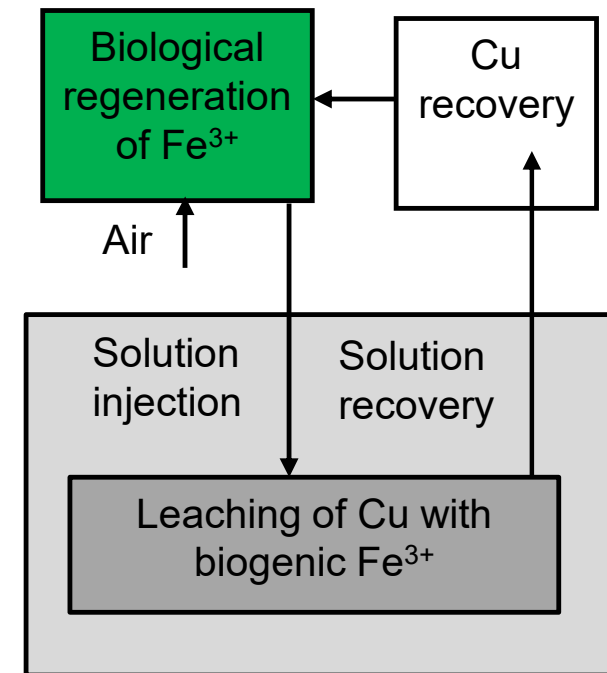
1. Water leaching



2. Acid leaching



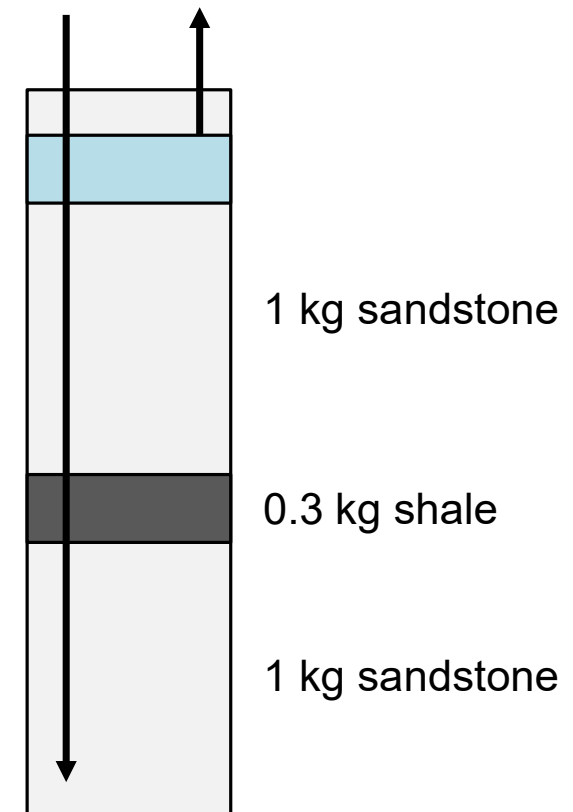
3. Copper leaching and recovery



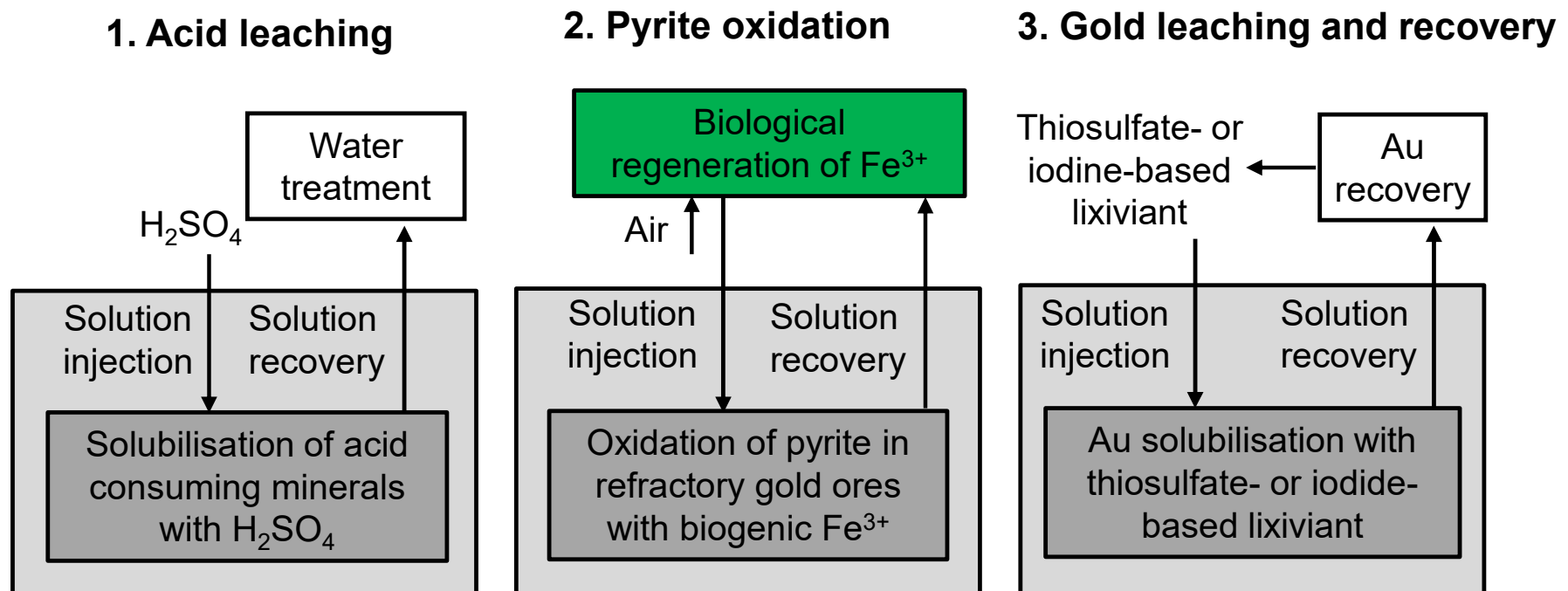
Adapted from: Kaksonen and Pedersen 2023. Chapter 17: The Future of Biomining: Towards Sustainability in a Metal-demanding World. In: Bryan, Johnson, Roberto and Schlömann (eds.) *Biomining Technologies: Bioprocessing Options for Extracting and Recovering Metals from Ores and Wastes*. Springer-Verlag (Heidelberg, Germany). Pp. 295-314.

Lab-scale column study for evaluating *in situ* biomining of copper from a saline calcareous copper sulfide ore

- Kupferschiefer comprised of sandstone (2.98% Cu) and black shale (3.61% Cu), with Cu mainly in chalcocite (Cu_2S), bornite (Cu_5FeS_4), chalcopyrite (CuFeS_2),
- Lab-scale experiment with upflow columns and acidophilic iron- and sulfur-oxidisers
- Three steps:
 - 19 d water leaching
 - 3 d acid leaching (0.9 M sulfuric acid)
 - 40 d bioleaching (Cu removed with sulfide precipitation 7 time from bleed stream)
- Leaching yield: 59% Cu



In situ leaching of gold from refractory sulfidic deposit after biological pre-treatment



Adapted from: Kaksonen and Pedersen 2023. Chapter 17: The Future of Biomining: Towards Sustainability in a Metal-demanding World. In: Bryan, Johnson, Roberto and Schlömann (eds.) *Biomining Technologies: Bioprocessing Options for Extracting and Recovering Metals from Ores and Wastes*. Springer-Verlag (Heidelberg, Germany). Pp. 295-314.

Column study to evaluate *in situ* biooxidation of Au-bearing pyritic ores before gold leaching

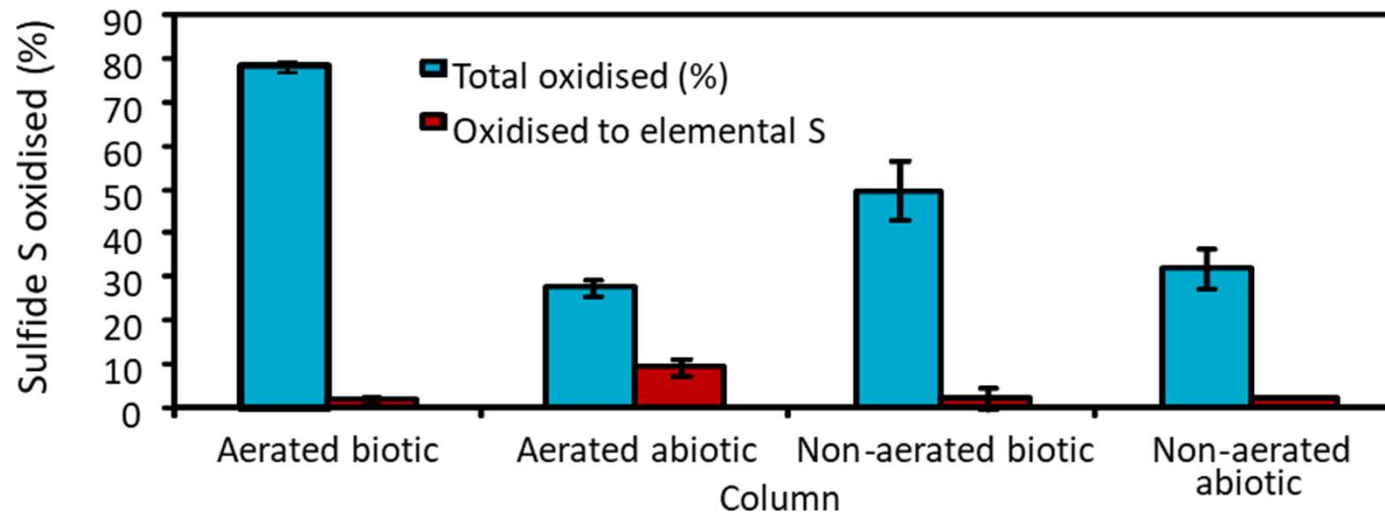


Upflow columns:

- 1.6 kg ore
- Particle size 1-4 mm
- S content 0.93%

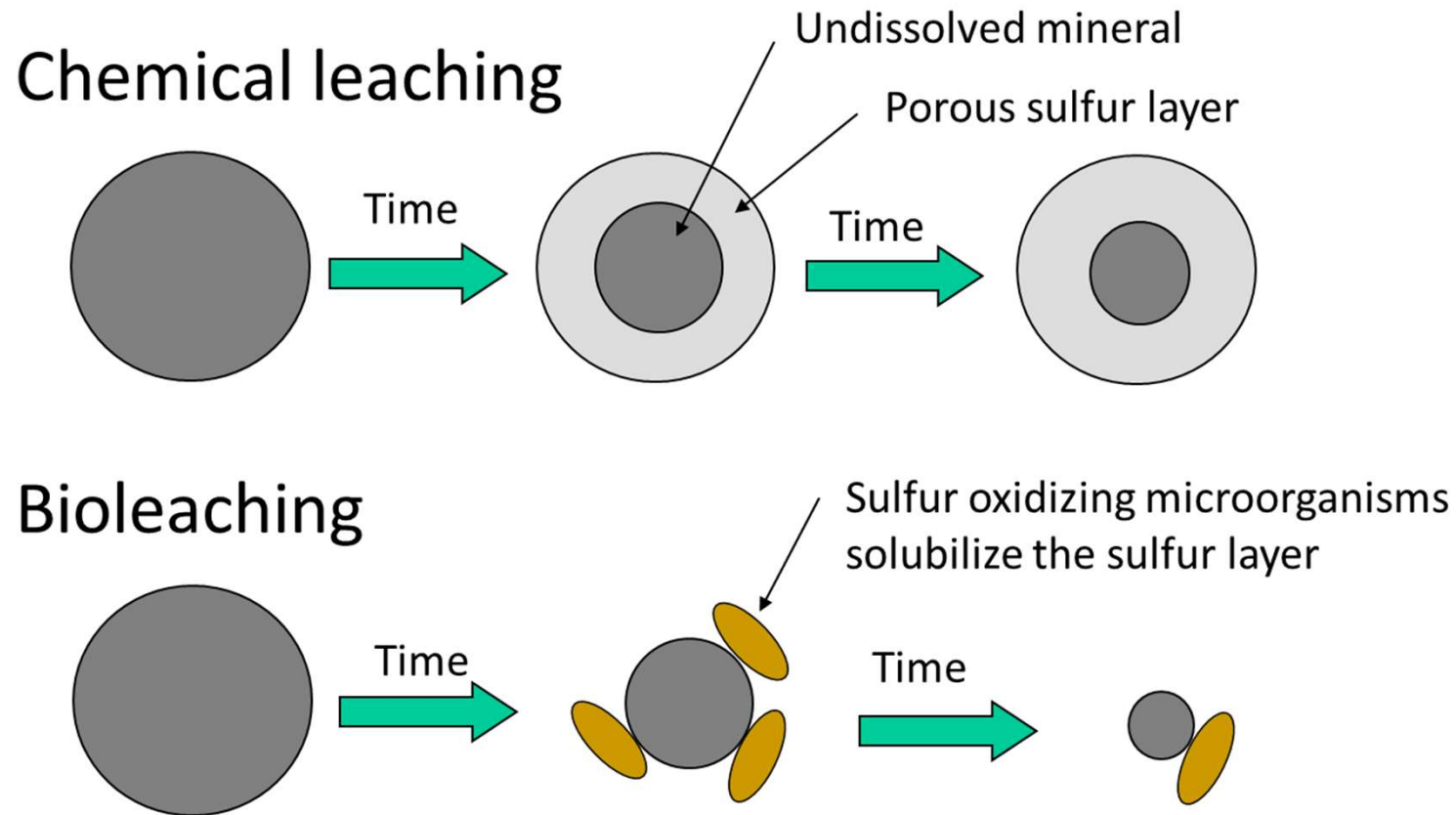
- Ore washed with pH 1.5 distilled H₂O to decrease acid consumption
- Biologically generated ferric sulfate (26 g Fe²⁺ L⁻¹ pH 1.7) pumped through 158 d
- Columns 1 and 2 aerated from the bottom at 0.6-0.8 L min⁻¹
- Thymol (0.4 g/L) added to abiotic (chemical control) column influents (2 and 4)

Key findings from the upflow column study



- Pyrite was oxidised under aerobic and anaerobic conditions using biologically generated ferric iron
- The simulated underground aeration and the presence of bioleaching microorganisms enhanced pyrite oxidation
- Microorganisms decreased the accumulation of S^0 , the presence of which may decrease subsequent gold recovery

Microbiological removal of passivating sulfur layers



Adapted from Crundwell FK. 2003. How do bacteria interact with minerals. Hydrometallurgy 71: 75-81.

Factors affecting bioleaching

Mineralogical factors:

- Mineral composition
- Liberation
- Porosity/permeability

Physical-chemical factors:

- Temperature
- Pressure
- Solubility of gases (O₂, CO₂)
- pH
- Redox
- Fe²⁺, Fe³⁺
- Other metals

Bioleaching



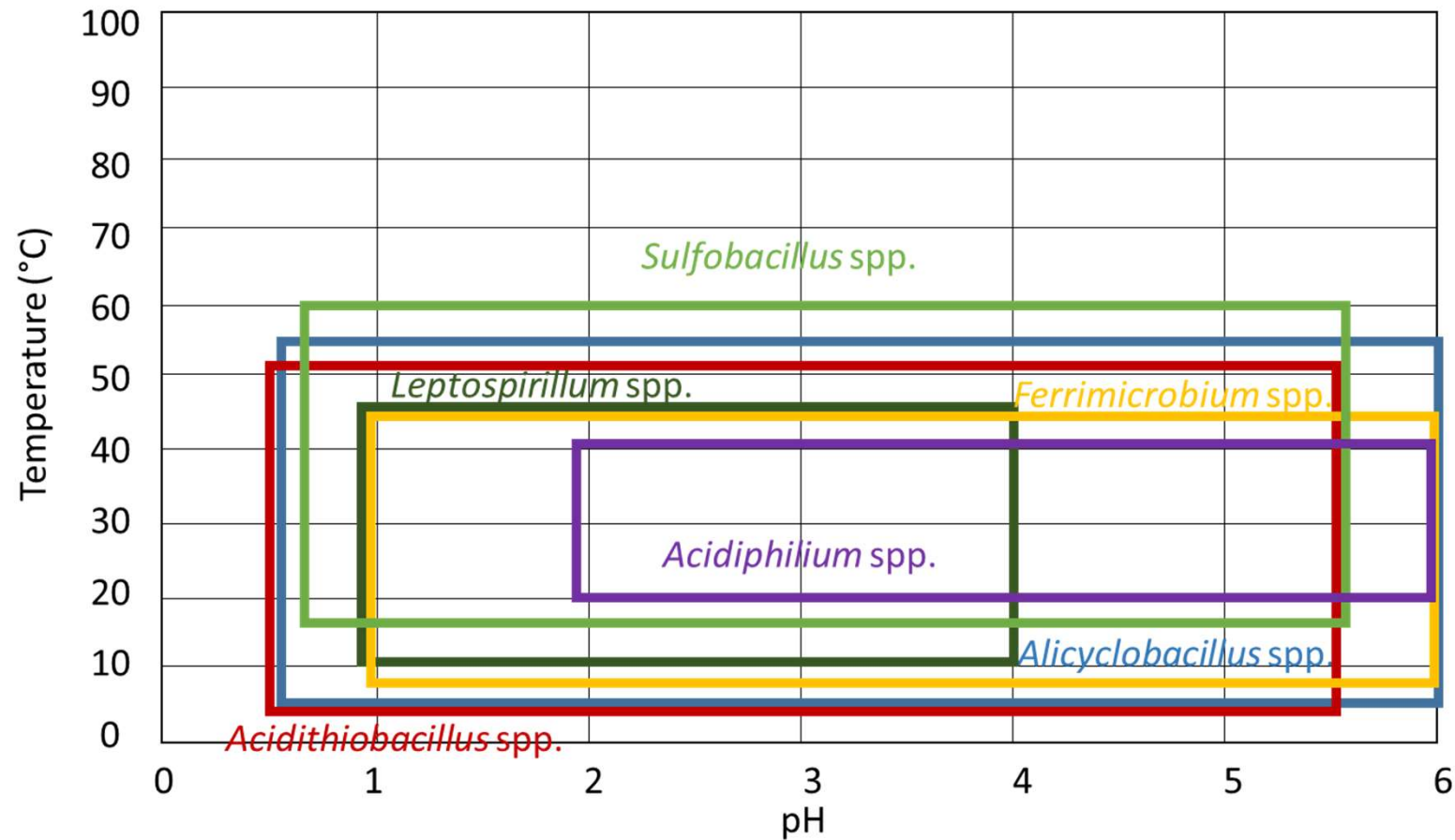
Process factors:

- Solid/liquid ratio
- Retention time

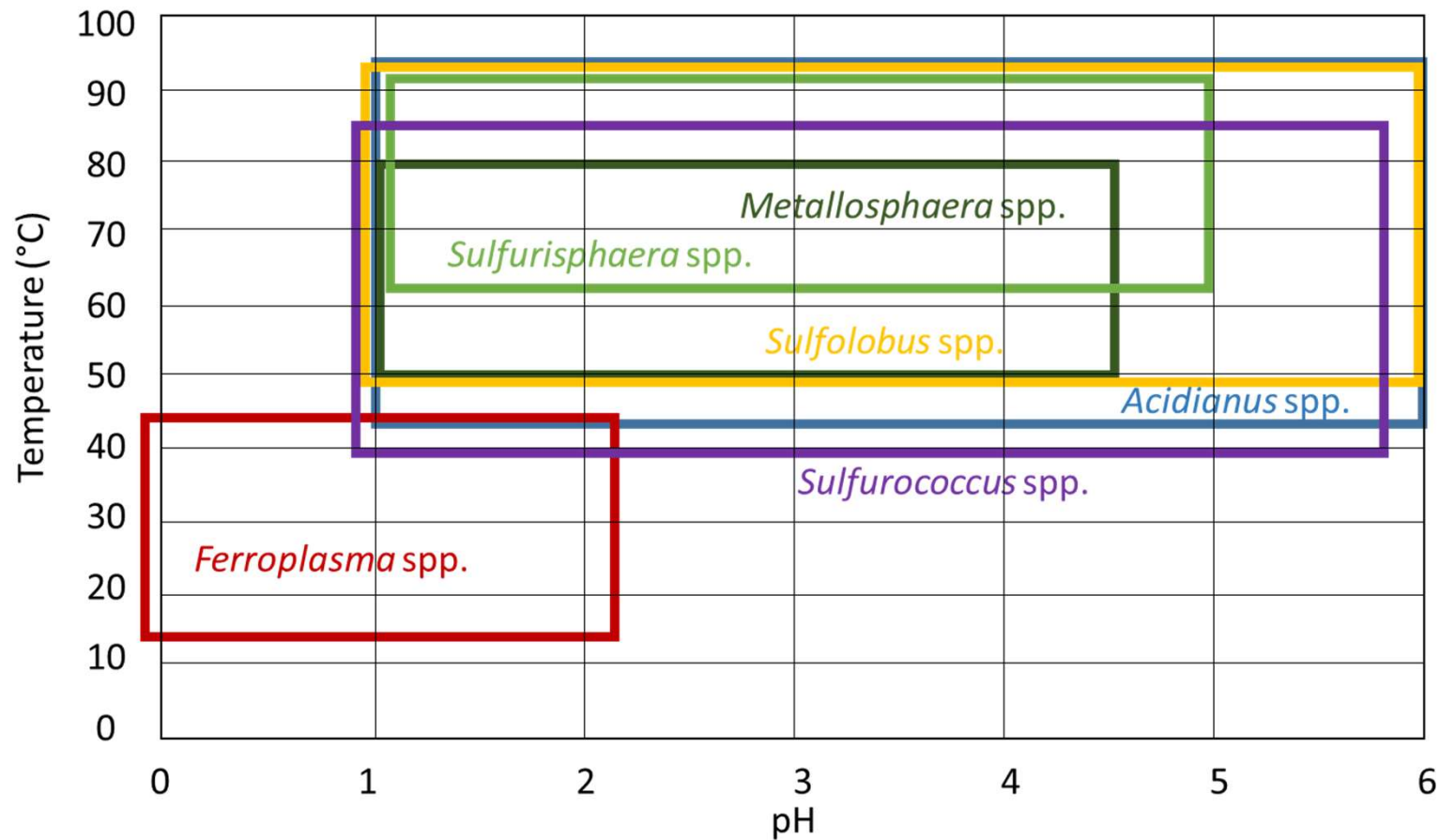
Microbiological factors:

- Strains
- Diversity
- Density
- Activity
- Distribution
- Substrate availability
- Tolerance

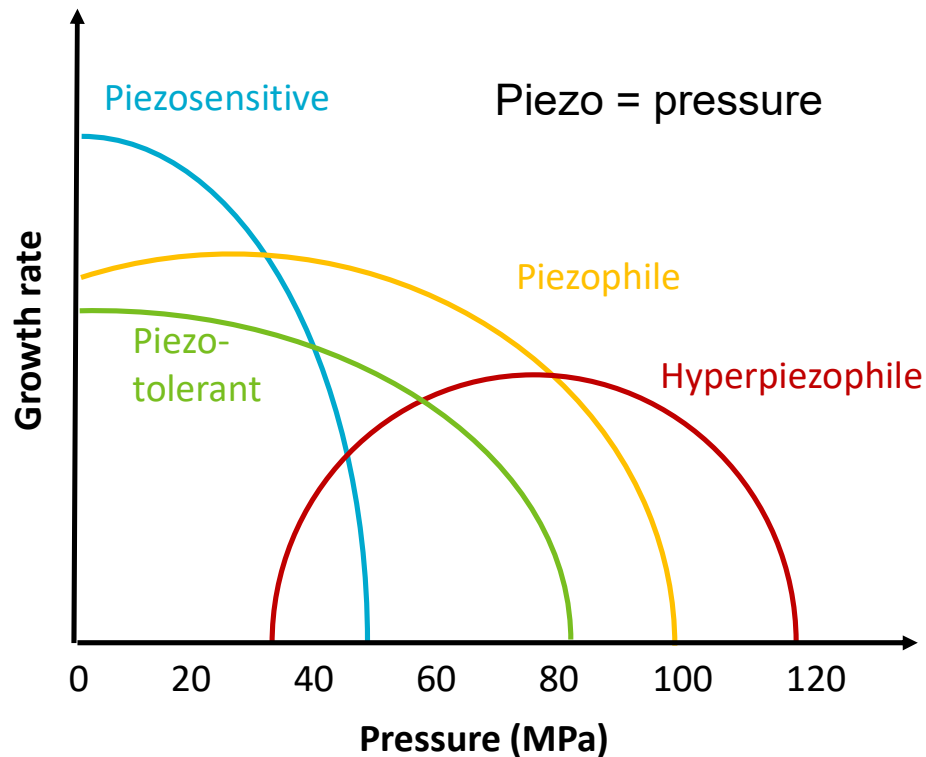
Temperature and pH ranges of bioleaching bacteria



Temperature and pH ranges of bioleaching archaea



Pressure requirements for microorganisms



Adapted from:

Abe F, Horikoshi K. 2001. The biotechnological potential of piezophiles. *Trends in Biotechnology* 19(3): 102-8.

Fang J, Zhang L, Bazylinski DA. 2010. Deep-sea piezosphere and piezophiles: geomicrobiology and biogeochemistry. *Trends in Microbiology* 18: 413-422.

- Pressure increases with increasing depth approximately 27.4 kPa km^{-1} in continental crust (Jones and Lineweaver, 2010b).
- Limit of liquid water on Earth at a depth of $\sim 75 \text{ km}$ ($p = 3,000 \text{ MPa}$ and $T = 431^\circ \text{C}$) \rightarrow depth limit for hydrometallurgical ISR (Jones and Lineweaver, 2010a).
- *Acidithiobacillus ferrooxidans* is tolerant to hydrostatic pressure of 15.2 MPa (Hiskey 1994; Torma 1975)
- Active and viable *Escherichia coli* and *Shewanella odeinensis* detected at pressures of up to $1,060 \text{ MPa}$ and $1,680 \text{ MPa}$, respectively, equivalent to depths of $\sim 35 \text{ km}$ and $\sim 50 \text{ km}$ below Earth's crust (Sharma et al., 2012)

Conclusions

- Bioleaching utilises acidolysis, redoxolysis, complexolysis
- Bioleaching mechanisms include contact, non-contact and collaborative bioleaching
- *In situ* bioleaching can be conducted under saturated or unsaturated conditions for a range of minerals and commodities
- Multiple factors impact biomining efficiency and can be optimised to improve leaching yields
- The application of biomining to ISR can enable resource extraction from low grade, complex and deep ore deposits, reduce passivation of mineral surfaces and environmental impacts of mining

Thank you

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